

TABLE I
 SUMMARY OF PROPERTIES OF NEW COMPOUNDS

Compound	B.P.		M.P., °C.	n_D^{20}	d_4^{20}	Molar Refraction		AR _F Calcd.
	°C.	Mm. Hg				Calcd. ^a	Found	
<i>n</i> -C ₃ F ₇ CH ₂ CH ₂ I	112-113	628	-5	1.3771	1.918	38.10	38.89	1.21
	77.8	185						
	44.3	47						
<i>n</i> -C ₃ F ₇ CH ₂ CH ₂ OCOCH ₃	132	633		1.3283	1.4135	35.49	36.8	1.24
<i>n</i> -C ₃ F ₇ CH ₂ CH ₂ OH	116-117	632		1.3151	1.506	26.83	27.81	1.24
	62-63	111						
<i>n</i> -C ₃ F ₇ CH ₂ COOH	170-172 ^b	630	10	1.3202	1.604	26.84	28.21	1.31
	62.3	6						
<i>n</i> -C ₃ F ₇ CH ₂ CONH ₂			92.5-93					
<i>n</i> -C ₃ F ₇ (CH ₂) ₂ CONH ₂			96.5-97					
<i>n</i> -C ₃ F ₇ (CH ₂) ₃ CONH ₂			102.5-103					
<i>n</i> -C ₃ F ₇ COCH ₂ Cl	97-98	625		1.3240	1.580	30.18	31.8	1.21
	61	178						
<i>n</i> -C ₃ F ₇ COCH ₂ Br	118	631		1.3436	1.818	33.08	33.88	1.22
	80.2	213						
C ₂ F ₅ COCH ₂ Cl	74	634		1.3088	1.348	26.55	27.8	1.34

^a Calculated from the Lorenz-Lorentz formula with 1.1 as AR_f for fluorine. ^b With dec.

Method II. Twenty-three grams (0.1 mole) of C₃F₇COCl dissolved in 150 ml. of sodium-dried ether was added dropwise to 500 ml. of ethereal diazomethane (prepared from 35 g. of *N*-methylnitrosourea) contained in a 1-l. three-neck flask fitted with a dropping funnel and stirrer. The third neck was protected with a calcium chloride drying tube. The reaction mixture was kept cold (0° to -10°) during the addition of the acid chloride.

When the addition was completed the mixture was allowed to warm to room temperature and allowed to stand for 8 hr. The dropping funnel was replaced by a gas inlet tube, and a bubbler was attached to the drying tube outlet. Anhydrous hydrogen chloride, passed successively through a safety trap, a concentrated sulfuric acid scrubber, and a second safety trap, was passed slowly into the solution. When the evolution of nitrogen ceased, the flow of hydrogen chloride was discontinued. The solution was then worked up as described in Method I. The yield of C₃F₇COCH₂Cl (b.p. 98.5-99.5°/632 mm.) was 7 g. (28.5%).

Preparation of heptafluoropropyl bromomethyl ketone. This reaction was carried out under conditions similar to that described for C₃F₇COCH₂Cl (Method I). Twenty-eight grams (0.1 mole) of C₃F₇COBr was used. The fraction boiling at 80-81° at 213 mm. was collected. The yield of α -bromoketone was 6 g. (20%), n_D^{20} 1.3436; d_4^{20} 1.818.

Anal. Calcd. for C₃H₂BrF₇O: C, 20.64; H, 0.69; Br, 27.46. Found: C, 20.69; H, 0.80; Br, 27.80.

Pentafluoroethyl chloromethyl ketone was prepared in a manner similar to that used for the preparation of C₃F₇COCH₂Cl.

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Action of Aluminum Chloride on Hexafluoropropene

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Aluminum chloride has been found to react with hexafluoropropene to yield CF₃CF=CFCl, CF₃CF=CCl₂, CF₂ClCF=CCl₂, CFCl₂CF=CCl₂, CCl₃CF=CCl₂ and CCl₃CCl=CCl₂. A mechanism for the reactions involving replacement as well as rearrangement is postulated.

The reaction of aluminum chloride with chloro-fluoroalkanes to replace fluorine by chlorine has been reported by Henne^{2,3} and Miller and his co-

workers.⁴⁻⁶ In the present work, the action of aluminum chloride on hexafluoropropene was studied to ascertain the order of replacement of the organically bound fluorine atoms by chlorine and the de-

(1) From the Ph.D. dissertation submitted to the University of Colorado, May 1953. E. I. du Pont de Nemours & Co., Inc., Pre-doctoral fellow: 1952-1953.

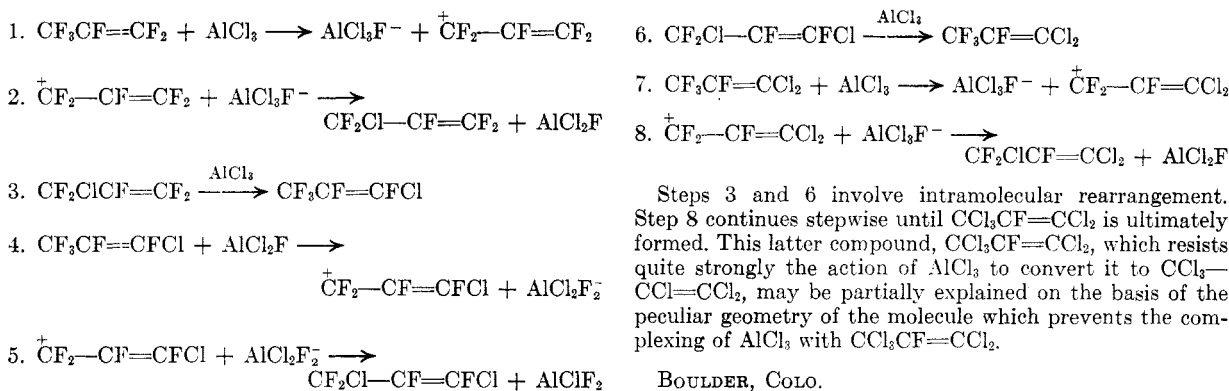
(2) A. L. Henne and H. M. Leicester, *J. Am. Chem. Soc.*, **60**, 864 (1938).

(3) A. L. Henne and M. S. Newman, *J. Am. Chem. Soc.*, **60**, 1697 (1938).

(4) W. T. Miller, Jr., *J. Am. Chem. Soc.*, **62**, 993 (1940).

(5) W. T. Miller, Jr., E. W. Fager, and P. H. Griswald, *J. Am. Chem. Soc.*, **72**, 705 (1950).

(6) W. A. Miller, Jr., U. S. Patent Application **47,553**. *Chem. Abstr.*, **47**, 4895 (1953).



Steps 3 and 6 involve intramolecular rearrangement. Step 8 continues stepwise until $\text{CCl}_3\text{CF}=\text{CCl}_2$ is ultimately formed. This latter compound, $\text{CCl}_3\text{CF}=\text{CCl}_2$, which resists quite strongly the action of AlCl_3 to convert it to $\text{CCl}_3-\text{CCl}=\text{CCl}_2$, may be partially explained on the basis of the peculiar geometry of the molecule which prevents the complexing of AlCl_3 with $\text{CCl}_3\text{CF}=\text{CCl}_2$.

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[CONTRIBUTION FROM THE LABORATORY FOR THE STUDY OF HEREDITARY AND METABOLIC DISORDERS, AND THE DEPARTMENTS OF BIOLOGICAL CHEMISTRY AND MEDICINE, UNIVERSITY OF UTAH]

Preparation and Properties of β -3-Indolyl Compounds Related to Tryptophan Metabolism¹

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3-Indolylpyruvic acid was prepared from DL-tryptophan *via* the *N*-chloroacetyl derivative and 2-methyl-4-(3'-indolal)-5-oxazolone, and also from 3-formylindole *via* 2-methyl-4-(1'-acetyl-3'-indolal)-5-oxazolone. The pyruvic acid was converted to β -(3-indolyl)lactic and β -(3-indolyl)- α -oximinopropionic acids. β -(3-Indolyl)acrylic and β -(3-indolal)malonic acids were synthesized from 3-formylindole and malonic acid. 3-Indolylglyoxylic acid, amide, and methyl ester were prepared from indole and oxalyl chloride *via* 3-indolylglyoxylyl chloride. 3-Indolylglycolic acid was obtained as a stable sodium salt by reduction of the glyoxylic acid and the instability of the free glycolic acid was confirmed. 3-Indolylcarboxylic acid was prepared from 3-cyanoindole which was obtained from 3-indolylglyoxylic acid or from 3-formylindole *via* the aldoxime. 3-Indolylacetamide was synthesized from 3-indolylacetic acid *via* the acid chloride. The factors which influence the yield, stability, and purity of these compounds are considered in relation to inadequacies in earlier literature.

Only a minor portion of the tryptophan ingested by man follows the known metabolic paths, which lead to nicotinic acid or to serotonin, and the fate of the remainder is uncertain.⁴ Varying small amounts of many indole compounds are present in human urine; an abnormal excretion of some of these compounds has been reported in cases of phenylketonuria,⁵ malignant carcinoid tumor,^{6,7} and Hartnup disease.⁸ The preparation of several 3-indolyl compounds, which were required in a study of urinary indole acids and their possible

significance in relation to other metabolic paths,⁹ is reported in the present paper. New syntheses of indolepyruvic acid, sodium indoleglycolate, and indoleacetamide are presented, together with effective procedures for indoleacrylic, indolecarboxylic, indoleglyoxylic, and indolelactic acids. The conditions which influence the yield, stability, and purity of the compounds are considered. These factors have not been treated sufficiently in many of the earlier publications, and procedures frequently have not been described or are inadequate.

β -(3-Indolyl)pyruvic acid (I) was prepared in 43% overall yield from DL-tryptophan *via* its *N*-chloroacetyl derivative (II) and 2-methyl 4-(3'-indolal)5-oxazolone (III). Cooley and Wood¹⁰ used this approach to make *N*-acetyldehydrotryptophan, but did not isolate I and III. I also was obtained in low yield *via* 2-methyl-4-(1'-acetyl-3'-indolal)-5-oxazolone by condensing 3-formylindole with acetylglycine. Bentley *et al.*¹¹ recently described the

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(4) For recent reviews on the metabolism of tryptophan, see A. H. Meister, *Biochemistry of the Amino Acids*, Academic Press Inc., New York, 1957, pp. 333, 407, and C. E. Dalglish, *Advances in Protein Chemistry*, **10**, 79 (1955).

(5) M. D. Armstrong and K. S. Robinson, *Arch. Biochem. Biophys.*, **52**, 287 (1954).

(6) A. Sjoerdsma, H. Weissbach, and S. Udenfriend, *Am. J. Med.*, **20**, 520 (1956).

(7) G. Curzon, *Arch. Biochem. Biophys.*, **66**, 497 (1957).

(8) J. B. Jepson, *Biochem. J.*, **64**, 14P (1956).

(9) M. D. Armstrong, K. N. F. Shaw, M. G. Gortatowski, and H. Singer, *J. Biol. Chem.*, **232**, 17 (1958).

(10) S. L. Cooley and J. L. Wood, *J. Biol. Chem.*, **185**, 287 (1950).

(11) J. A. Bentley, K. R. Farrer, S. Housley, G. F. Smith, and W. C. Taylor, *Biochem. J.*, **64**, 44 (1956).